

ASYMMETRIC SYNTHESIS OF (S)-METHYL-3-HYDROXYALKANOATES  
FROM KETENE AND 2,2-DICHLOROALDEHYDES VIA  
4-(1,1-DICHLOROALKYL)-2-OXETANONES

Peter E.F. Ketelaar, Eiel G.J. Staring and Hans Wynberg\*  
Department of Organic Chemistry, University of Groningen,  
Nijenborgh 15, 9747 AG Groningen, The Netherlands

Abstract: Using the quinidine catalyzed cycloaddition reaction of ketene and 2,2-dichloroaldehydes, the preparation of several optically pure (S)-methyl-3-hydroxyalkanoates is reported.

Recently we reported a reasonably efficient enzymic synthesis of 1-octanol and optically active 1,2-epoxyoctane (70% e.e.) from n-octane and 1-octene by *Pseudomonas oleovorans*.<sup>1,2,3</sup>

We had reasons to believe that the intracellular inclusions<sup>4</sup> formed during this oxidation contained poly-3-hydroxyoctanoate.<sup>5</sup> The absolute configuration of the chiral centers of poly-3-hydroxybutanoate, also a well known constituent of intracellular inclusions<sup>6</sup> can be correlated to methyl-3-hydroxybutanoate by methanolysis of the polymer and was shown to be R.<sup>7</sup> In the same way the R-configuration was assigned to the chiral centers in poly-3-hydroxypentanoate.<sup>8</sup> In order to be able to confirm the structure of the monomer and to assign the absolute configuration of the chiral centers in poly-3-hydroxyoctanoate, we developed an independent synthesis of optically pure methyl-3-hydroxyalkanoates. Encouraged by the success of our synthesis of (S)-methyl-3-hydroxybutanoate from (R)-4-(trichloromethyl)-2-oxetanone,<sup>10,11</sup> obtained from the quinidine catalyzed cycloaddition of chloral and ketene, we decided to develop a synthesis of (S)-methyl-3-hydroxyalkanoates, using (R)-4-(1,1-dichloroalkyl)-2-oxetanones (3a,b,c) as chiral educt. The sequence of reactions leading to the desired products is depicted in Scheme I.

A typical procedure for the synthesis of these optically pure methyl-3-hydroxyalkanoates follows: 2,2-Dichloroaldehydes (2a,b,c) were prepared by direct chlorination according to the method described by de Buyck.<sup>9</sup> The crucial step in the reaction sequence is the asymmetric synthesis of (R)-4-(1,1-dichloroalkyl)-2-oxetanones (3a,b,c).<sup>15</sup> The formation of the chiral center was achieved by the reaction of the 2,2-dichloroaldehydes (2a,b,c) with ketene



scale, using Pd/C (10%) as a catalyst under a hydrogen atmosphere (3 atm. H<sub>2</sub>-pressure) in dry methanol as a solvent. It was necessary to add base (K<sub>2</sub>CO<sub>3</sub>, slight excess) to neutralize HCl formed during the reaction. Addition of a drying agent (MgSO<sub>4</sub>) proved to be advantageous. Probably the water liberated upon neutralization of HCl tends to hydrolyse the esters, thus decreasing yields. Flash chromatography (SiO<sub>2</sub>, pentane-ether, 1:3) and bulb to bulb distillation (60°C, 0.05 mmHg), yielded the esters,<sup>16</sup> (S)-methyl-3-hydroxyhexanoate (5a), c.y. 85%, [ $\alpha$ ]<sub>578</sub><sup>20</sup> 18.9, c 1, cyclohexane; (S)-methyl-3-hydroxy octanoate (5b), c.y. 87%, [ $\alpha$ ]<sub>578</sub><sup>20</sup> 23.6, c1, cyclohexane; (S)-methyl-3-hydroxy decanoate (5c), cy 83%, [ $\alpha$ ]<sub>578</sub><sup>20</sup> 25.8, c 1, cyclohexane.

No inversion takes place during hydrogenolysis, but change in R/S nomenclature is caused by change in substituents. Using the method by Feringa, Smaardijk and Wynberg,<sup>14</sup> the esters 5b and 5c were shown to have enantiomeric purities of >98%. Compound 5a is only 92% enantiomerically pure.

In this paper only the synthesis of (S)-methyl-3-hydroxyalkanoates is described. However, using optically pure (S)-2-oxetanones as chiral starting materials, allows the preparation of the (R)-enantiomers of the methyl-3-hydroxyalkanoates. The (S)-4-(1,1-dichloroalkyl)-2-oxetanones can be prepared using quinine or benzoylquinine as a catalyst in the cycloaddition reaction of ketene and the dichloroaldehydes.

Physical, analytical and spectroscopic data of compounds 5a,b,c are as follows:

5a. Methyl-3-hydroxy hexanoate: C<sub>7</sub>H<sub>14</sub>O<sub>3</sub> (MW = 146); b.p. 55-58°C, 0.05 mmHg. Anal. calcd.: C, 57.51; H, 9.65; O, 32.84. Found: C, 58.10; H, 9.85; O, 32.55. MS: M<sup>+</sup> at m/e = 145; IR: 1110, 1160, 1400, 1430, 1440, 1720, 2865, 2930, 2975, 3480 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 60 MHz):  $\delta$  0.94 ppm (m, 3H), 1.46 (m, 4H), 2.45 (d, 2H), 3.36 (s, 1H), 3.72 (s, 3H), 3.99 (t, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50.32 MHz):  $\delta$  13.7 (q), 18.4 (t), 38.5 (t), 41.0 (t), 51.4 (q), 67.5 (d), 173.1 (s).

5b. Methyl-3-hydroxyoctanoate: C<sub>9</sub>H<sub>18</sub>O<sub>3</sub> (MW = 174); b.p. 63-65°C, 0.05 mmHg. Anal. calcd.: C, 62.04; H, 10.41; O, 27.55; found: C, 61.11, H, 10.37; O, 28.22. MS: M<sup>+</sup> at m/e = 173; IR: 1120, 1160, 1410, 1440, 1450, 1730, 2860, 2925, 2970, 3520; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 60 MHz):  $\delta$  0.89 (m, 5H), 1.40 (m, 6H), 2.43 (d, 2H), 3.37 (s, 1H), 3.68 (s, 3H), 3.98 (t, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50.32 MHz):  $\delta$  13.8 (q), 22.4 (t), 25.0 (t), 31.5 (t), 36.4 (t), 41.1 (t), 51.5 (q), 62.9 (d), 173.2 (s).

5c. Methyl-3-hydroxydecanoate: C<sub>11</sub>H<sub>22</sub>O<sub>3</sub> (MW = 202); b.p. 74-76°C, 0.05 mmHg. Anal. calcd.: C, 65.35; H, 10.96; O, 23.64; found: C, 64.95; H, 10.64; O, 24.14. MS: M<sup>+</sup> at m/e = 201; IR: 1125, 1170, 1410, 1440, 1460, 1730, 2850, 2920, 2970, 3470; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 60 MHz):  $\delta$  0.88 (m, 6H), 1.39 (m, 9H), 2.45 (d, 2H), 3.35 (s, 1H), 3.70 (s, 3H), 3.95 (t, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50.32 MHz):  $\delta$  13.9 (q), 22.5 (t), 25.3 (t), 29.0 (t), 29.3 (t), 31.6 (t), 36.3 (t), 41.0 (t), 51.5 (q), 67.8 (d), 173.3 (s).

References

1. De Smet, M.J.; Kingma, J.; Witholt, B. and Wynberg, H., Enzym. Microbiol. Technol. 1983, 5, 352.
2. De Smet, M.J.; Wynberg, H. and Witholt, B., Applied and Environmental Microbiology 1981, 42, 811.
3. De Smet, M.J.; Witholt, B. and Wynberg, H., J. Org. Chem., 1981, 46, 3128.
4. De Smet, M.J., Thesis, Groningen, 1982.
5. De Smet, M.J.; Eggink, G.; Witholt, B.; Kingma, J. and Wynberg, H., J. of Bacteriol. 1983, 154, 870.
6. Shively, J.M., Ann. Rev. Microbiol. 1974, 28, 167.
7. Seebach, D. and Zuger, M., Helv. Chim. Acta 1982, 65, 495.
8. Seebach, D.; Brienne, M.J.; Renaud, Ph., Schweizer, W.B.; and Zuger, M., Helv. Chim. Acta 1984, 67, 1843.
9. De Buyck, L.; Veske, R.; de Kimpe, N.; Cantheyn, D. and Schamp, N., Bull. Soc. Chim. Belg. 1980, 89, 441.
10. Wynberg, H. and Staring, E.G.J., J. Am. Chem. Soc. 1982, 104, 166.
11. Wynberg, H. and Staring, E.G.J., J. Org. Chem., 1985, 50, 1977.
12. Jacques, J.; Collet, A. and Wilen, S.H., "Enantiomers, Racemates and Resolutions", 1st ed., John Wiley & Sons N.Y., 1981.
13. Wynberg, H. and Staring, E.G.J., J. Chem. Soc., Chem. Commun. 1984, 1181.
14. Feringa, B.L.; Smaardijk, A.A. and Wynberg, H., J. Am. Chem. Soc., accepted for publication.
15. Patent pending: Wynberg, H.; Staring, E.G.J., PCT/NL 83/00040.

(Received in UK 16 July 1985)